

# Conventional and alternative treatment approaches for *Clostridium difficile* infection

#### Khalid M. Aljarallah

Department of Medical Laboratory Sciences, College of Applied Medical Sciences, Majmaah University, Majmaah, KSA

Address for correspondence: Dr. Khalid M. Aljarallah, Department of Medical Laboratory Sciences, College of Applied Medical Sciences, Majmaah University, Majmaah, KSA. E-mail: K.aljarallah@mu.edu.sa

ijhs.org.sa

1658-3639

PUBLISHER: Qassim University

AD	CTT1		$\mathbf{C}$
AD		NA	

*Clostridium difficile*-associated disease continues to be one of the leading health concerns worldwide. *C. difficile* is considered as a causative agent of nosocomial diarrhea that causes serious infection, which may result in death. The incidences of *C. difficile* infection (CDI) in developed countries have become increasingly high which may be attributed to the emergence of newer epidemic strains, extensive use of antibiotics, and limited alternative therapies. The available treatment options against CDI are expensive and promote resistance. Therefore, there is urgent need for new approaches to meet these challenges. This review discusses the current understanding of CDI, the existing clinical treatment strategies and future potential options as antidifficile agents based on the available published works.

Keywords: Alternative treatment, Clostridium difficile, Clostridium difficile infection

Introduction

WEBSITE:

ISSN:

*Clostridium difficile* is an anaerobic, spore-forming, Grampositive bacterium, the most common pathogen that can cause antibiotic-associated diarrhea. Normal gut flora usually resists colonization and overgrowth of *C. difficile*.<sup>1</sup> *C. difficile* infection (CDI) represents a serious challenge to medical practice in many developed countries, however, information from developing countries on this infection is generally lacking. One report on CDI in China indicated that such infection was present but no systematic study was conducted to provide more complete epidemiology data.<sup>2</sup> One reason is the lack of affordable diagnostic kits to be used for clinical or epidemiology studies.

The uses of antibiotics usually modify the intestinal microbiome and permit propagation of C. difficile. It has been noticed that hospitalized patients are the prime target of CDI, although C. difficile also present as a colonizer in 2-3% of healthy individuals and 70% in healthy children.<sup>3</sup> CDI is associated with antibiotic treatments and could occur in every kind of clinical setting. Recent reports have shown that 20% of individuals who were hospitalized become colonized with C. difficile during hospitalization, and about 30% of those patients can develop diarrhea.<sup>4</sup> Although CDI is commonly regarded as mild-to-moderate diarrheal disease with abdominal pain, in severe cases CDI can present with acute abdomen as well as fulminant and life-threatening colitis. The diagnosis of C. difficile colitis should be suspected in any patient with diarrhea, who has been under antibiotics treatment over the previous 3 months or has been recently hospitalized.<sup>5</sup> However, it has been recently reported that C. difficile can be a cause of diarrhea in community without any previous hospitalization or antibiotic exposure.<sup>6</sup> *C. difficile* produces heat-resistant spores that can persist in the environment for several months, thus providing the basis for nosocomial outbreaks even after extensive cleaning measures.

#### Pathophysiology

Pathogenic strains of C. difficile produce multiple diverse toxins.7 The preeminent categorized toxins are toxin A, which is an enterotoxin, and toxin B, a cytotoxin. Both toxins are of high molecular weight proteins (308 and 270 kDa, respectively) having the capability of binding to some specific receptors on intestinal mucosal cells of the host. Receptor-bound toxins usually gain their intracellular entry using catalyst specific Rho proteins that support in actin polymerization, cytoskeletal rearrangements and cell movement.8,9 Both toxin A and B emerge to play an essential role in the pathogenesis of C. difficile colitis in humans through the initiation of apoptosis in the target mucosal cells. Toxin damage of the colonic mucosa always escort to an accumulation of fibrin, mucin, and dead cells, finally structuring a layer of debris in the colon. Subsequent inflammatory activation adjoins to the direct toxin-associated damage resulting in mild diarrheal disease up to extensive intestinal wall damage with septic shock and death.7,10

## **Clinical Symptoms**

It is important and crucial to differentiate between asymptomatic colonization and symptomatic CDI. Individuals who are

colonized by the organism may attain protection against development of the infection but they stay as a potential transmission source of the disease in health-care settings.<sup>11</sup> Symptom starts from simple irritation of mucosa, watery to soft diarrhea with a sweetish, and foul odor<sup>12</sup> to the full clinical symptom of pseudomembranous colitis with typical endoscopic findings, preferentially in the region of the sigmoid and rectum. CDI affects the right colon alone is rarer.<sup>13</sup> Furthermore, stool frequency of the patient may be increased  $\times 10$ /day. In older patients, the signs of toxicosis requiring treatment can take place rapidly. If such symptoms prolonged, hypoalbuminemia, and protein-losing enteropathy can occur.<sup>14</sup> Subfebrile temperatures are common.<sup>15</sup> On physical examination, the colon is swollen in the lower left abdomen. There is usually slight local pain on palpation.<sup>12,13</sup> Aggressive signs of complicated CDIs with ileus, toxic megacolon, perforation, or sepsis (<5% of cases) include absence of colonic peristalsis, sudden-onset constipation, extreme leukocytosis, and high fever.<sup>12-15</sup> Moreover, further diagnostic measures may be required, i.e., contrast computed tomography of the abdomen.<sup>13</sup> Different professional societies of gastroenterologists as well as infectious disease specialists have recently published upto-date guidelines for CDI management.<sup>16-18</sup> The European Society of Clinical Microbiology and Infectious Diseases have recently published guidelines in 2014.18 The definition of an event of CDI comprises a clinical picture (Table 1) compatible with CDI, microbiological evidence of free toxins, and the presence of the organism in stool without realistic evidence of another cause of diarrhea, or pseudomembranous colitis (diagnosed by endoscopy, after colectomy, or on autopsy).

**Table 1:** Patient characteristics correlating with disease severity

 when associated with CDI (adapted from Debast *et al.*, [2014])

Category	Signals/Symptoms
Physical examination	Fever, rigors, hemodynamic instability including signs of shock, respiratory failure with need for mechanical ventilation, peritonism, ileus
Laboratory tests	Leukocytosis>15 Gpt/L, left shift with>20% neutrophils, rise in serum creatinine>1.5×baseline, lactate>5 mmol/L, albumin<30 g/L
Endoscopy imaging	Presence of pseudomembranes colonic distension>6 cm in transverse colon/toxic megacolon, colonic wall thickening, pericolonic fat stranding, ascites due to CDI

CDI: Clostridium difficile infection

#### Diagnosis

The international CDI diagnosis guidelines recommend evidence-based, rapid detection of toxigenic CDI from stool samples.<sup>16-22</sup> Multistep diagnostic procedures are recommended (Table 2).<sup>23</sup>

Only the symptomatic patients should be tested. Repeated stool samples are not usually required. Rapid antigen tests and nucleic acid amplification tests are mostly important in routine diagnosis, due to their short turnaround time (TAT), ranges from 15 min to 3 h. The toxigenic culture, i.e., the anaerobic culture in a special media, combined with evidence of the toxin in the culture supernatant, is considered as the diagnostic gold standard. Anaerobic culture is required for further antibiotic resistance and ribotype testing. Cultures are not well suited for acute diagnosis, as they have a long TAT (>72 h). A macroscopic finding of pseudomembranous colitis is in many cases so characteristic that CDI can also be diagnosed via endoscopy or colonoscopy, though with limited sensitivity.

#### C. difficile therapy

Having a confirmed CDI, it is crucial that proper infection control measures are in place to avoid further spread of the infection within the same ward or hospital. To avoid spreading of *Clostridium* spores, hands need to be washed, patients should kept in isolation, and importantly, gloves and protective clothing must be worn by all staff along with continuous hand hygiene after each patient contact.<sup>24</sup> In all patients with CDI, it is necessary to stop the causing antibiotic therapy. This may be suitable as the only treatment in a patient with little symptoms. Patients need appropriate replacement and monitoring of fluids and electrolytes, and antimotility drugs should be avoided. Currently, all recommendations for antibiotic therapy are based on differentiation between mild-to-moderate or severe disease.<sup>18</sup> Other guidelines further differentiate a severe to a complicated course.<sup>16-18</sup> For successful treatment of CDI, it is important for clinicians to start screening for risk factors of severe disease and parameters linked with an unfavorable outcome. There are many clinical and laboratory variables that correlate with severity of outcome as already explained in Table 1.18 In general, the most important prognostic indicators of severe

Fable 2: I	Microbiological	diagnostic tes	sts for C.	difficile and	their value (Adopte	ed from Lübbert et	al., [2014])
------------	-----------------	----------------	------------	---------------	---------------------	--------------------	--------------

Test	Indication
Test GDH EIA (TAT<2 h)	Initial screening test with high sensitivity and high negative predictive value; GDH-positive samples must undergo a confirmation test for the toxigenic infection
Toxin A and B EIA (TAT<2 h)	Confirmation test for toxigenic infection in GDH-positive samples (two-step algorithm); good correlation with severe infections, limited sensitivity; NAAT (three-step algorithm) recommended if no toxin detected
CTNA (TAT<24 h)	Standard test for evidence of toxins in stool; CTNA is rarely used for routine diagnosis, however, due to its longer TAT and low potential for standardization and automation
NAAT of toxin genes (TAT<4 h)	Confirmation test for toxigenic infection. NAAT (e.g., PCR) not recommended as screening test, as asymptomatic <i>C. difficile</i> carriers not requiring treatment or isolation may also be detected.

GDH: Glutamate dehydrogenase, CTNA: Cytotoxin neutralization assay, TAT: Turn-around time, EIA: Enzyme immunoassay, NAAT: Nucleic acid amplification test, PCR: Polymerase chain reaction, C. difficile: Clostridium difficile, CDI: Clostridium difficile infection

disease are; the age of >65 years, leukocytosis (>15 Gpt/L), decreased serum albumin (<3 g/L), rise in serum creatinine (>133 µmol/L or >1.5 times of the premorbid level) and underlying comorbidities.<sup>25-27</sup> In a patient with strong suspicion of CDI, empirical treatment is required according to European and American guidelines. This approach is only recommended for the patients with severe disease and/or risk factors for an unfavorable outcome. Furthermore, cessation of excessive antibiotic treatment is compulsory.

#### Antibiotics

Recent advances in the CDI antibiotic resistance have been reviewed by Spigaglia.<sup>28,29</sup> However, the following antibiotics are well known for the treatment of CDI and received a considerable attention by different researchers.

#### Metronidazole (MTZ)

MTZ is considered as the first choice for mild-to-moderate CDI.<sup>30</sup> Although the percentage of *C. difficile* strains resistant to MTZ is, in general, low, several studies have emphasized treatment failure with MTZ.<sup>31</sup> An elevated linear mean of minimum inhibitory concentrations (MICs) to MTZ have recently been observed in different strains; RT027 (1.1-1.42 mg/L), RT001/072 (0.65 mg/L), RT106 (0.65 mg/L), RT356 (0.61 mg/L) and in the nontoxigenic RT010 (1.5 mg/L), compared with the values of the other RTs (0.13-0.41 mg/L).<sup>32</sup> A study in Saudi Arabia by Alqumber reports susceptibility of MTZ with MIC range between 3 and 8  $\mu$ g/ml.<sup>33</sup> Another study, however, reports a spread of strains RT027 with reduced susceptibility to MTZ in Jerusalem, where they cause severe infections and a wide outbreak in 2013.<sup>34</sup>

Detection of strains with reduced susceptibility to MTZ can be challenging. This resistance is often unstable and laboratory manipulation of strains frequently results in MIC decrease toward a susceptibility range.35 Experimental methodology may affect the magnitude of measured MTZ MICs for C. difficile. The overall data reported in a recent study suggest the Agar Incorporation Method (AIM)<sup>36</sup> as the method of choice to detect strains with reduced susceptibility to MTZ compared with the Etest and the agar dilution method (AD).<sup>37</sup> Differences in the media used (Schaedlers broth and Wilkins-Chalgren agar for AIM and Brucella broth/agar for both Etest and AD) and in the duration of the precultured period (24 h for AIM and 48 h for both Etest and AD) seem to affect MIC determination.<sup>37,38</sup> MTZ susceptibility breakpoint for C. difficile defined by the CLSI and the European Committee on Antimicrobial Susceptibility Testing are not equivalent: The first is defined as 32 mg/L, the second >2 mg/L. Methodological variations and different interpretation categories may result in discrepancies, impacting therapeutic decision and comparison of data. Therefore, international committees are currently cooperating with the intention of harmonizing susceptibility testing and international breakpoints.

#### Vancomycin (VAN)

VAN, the first-line antibiotic treatment of choice for moderate to severe CDI,<sup>18,30</sup> consists of a glycosylated hexapeptide chain and cross-linked aromatic rings by aryl ether bonds, with a reduced absorption in the gastrointestinal tract.<sup>39</sup> Its mechanism of action results in inhibition of the biosynthesis of peptidoglycan, an essential component of the bacterial cell wall envelope.<sup>40</sup> Resistance to VAN has frequently observed in *Enterococci* and *Staphylococci*, but it is not so largely diffused in *C. difficile*. Although, a number of *C. difficile* strains with reduced susceptibility to VAN (MICs range >2-16 mg/L) have recently been described.

The mechanism of resistance in C. difficile is still unclear. Several Tn1549-like elements have been found in C. difficile.41,42 Differently from the original Tn1549 element described in Enterococcus faecalis, the Tn1549-like elements of C. difficile do not have a functional vanB operon. Recently, a vanG-like gene cluster, homologous to the cluster found in E. faecalis, have been described in a number of C. difficile isolates but, although this cluster is expressed, it is not able to promote resistance to VAN.43,44 MurG converts lipid I to lipid II during the membrane-bound stage of peptidoglycan biosynthesis. Alterations in this pathway, in VAN-resistant mutants, may affect VAN activity since VAN inhibits cell wall formation by binding to the D-Ala-D-Ala portion of lipid II.<sup>45</sup> Biofilm formation could be also involved in VAN-resistance. C. difficile within biofilms have been found to be more resistant to elevated concentrations of VAN (20 mg/L), and biofilm formation seems to be induced in the presence of subinhibitory and inhibitory concentrations of the antibiotic.46 The clinical significance of reduced susceptibility to VAN remains to be determined, since the fecal concentration of this antibiotic is very high, ranging between 520 and 2200 mg/L.47

Recommended treatment with MTZ and VAN according to disease severity is summarized in Table 3.

#### **Rifamycins and fidaxomicin (FDX)**

An increased rate of treatment failure and recurrence of infection have been associated with MTZ and VAN treatment,<sup>31</sup> therefore, other therapy options for CDI have been proposed in the recent years. Rifamycins, in particular rifaximin (RFX), have recently been prosed as "chaser therapy" for the treatment of relapsing CDI,<sup>48</sup> while FDX is a bactericidal new narrow spectrum macrocyclic antibiotic that is used for the management of CDI with high risk for recurrences.<sup>49</sup> Both rifampins (RIFs) and FDX are inhibitors of bacterial transcription but they have different RNA polymerase (RNAP) target sites. FDX binds to the "switch region" of RNAP, a target site that is adjacent to the RIF target but does not overlap.<sup>50,51</sup>

Susceptibility to RIF by either Etest or AD correlated completely with susceptibility to RFX.<sup>52</sup> Thus, testing susceptibility to RIF, a rifamycin that is related to RFX,

Severity	Criteria	Treatment	Comment
Mild-to-moderate disease	Diarrhea, no signs or symptoms of severe disease	MTZ 500 mg p.o. $3\times/day$ for 10 days	If no improvement in 5-7 days switch to VAN 4×125 mg p.o.
Severe disease	Two of the following: Albumin<30 g/L; leukocytosis>15 Gpt/L; creatinine>133 µmol/L; age>65 years; abdominal tenderness; comorbidities	VAN 125 mg p.o. 4×/day for 10 days	Other authors consider age<65 years and a rise in creatinine>1.5×baseline as equal risk factors for severe disease
Severe and complicated disease	Any of the following attributable to CDI: Admission to ICU for CDI; prolonged hypotension; ileus or significant abdominal distension; mental status changes; leukocytes>35 Gpt/L or<2 Gpt/L	VAN 500 mg p.o. 4×/day and MTZ 500 mg i.v. 3×/day and VAN per rectum (500 mg VAN in 500 ml Nalco 0.9%) 2-4×/day	Consider surgical consultation

Table	3.	Summary o	of recommended	treatment with	MTZ and	VAN :	according to	disease	severity (	Adant	ted from	Surawicz	et al	[2013])
Table	5.	Summary (	Ji recommended	i treatment with	IVI I Z allu	VAIN	according to	uisease	Severity (	Auap	ieu nom	Surawicz	zı uı.,	[2015])

ICU: Intensive care unit, CDI: Clostridium difficile infection, VAN: Vancomycin, MTZ: Metronidazole

can assess rifamycin class susceptibility in *C. difficile*. Data concluded from recent studies show that 11% of *C. difficile* clinical isolates are resistant to RIF and the rate of overall resistance appear to be rising.<sup>53-55</sup> *C. difficile* clinical isolates resistant to RIF have been detected in 17 out of 22 countries participating in a recent pan European surveillance and, in particular, high percentages of resistance (between 57% and 64%) have been observed in Italy, Czech Republic, Denmark and Hungary.<sup>32</sup> Prior exposure to RIFs has been reported to be a risk factor for RIF-resistant *C. difficile*<sup>52,56</sup> and resistant *C. difficile* strains may emerge even during therapy.<sup>57,58</sup> RIFs are commonly used as antituberculosis (TB) agents. Interestingly, all strains belonging to the emergent RT046 isolated in Poland from patients affected by TB and with a prolonged RIF therapy have been found highly resistant to these antibiotics.<sup>59</sup>

#### **Other antibiotics**

#### Tetracycline (TET)

Recent papers on the C. difficile resistance to TET show different results according to the countries where the studies took place. The resistance ranges between 2.4% and 41.67%.60-62 In this pathogen, resistance is commonly due to protection of the ribosomes from the action of antibiotic. The most widespread TET class in C. difficile is tetM, usually found on conjugative Tn916-like elements.<sup>63-65</sup> The Tn916-like family is responsible for the spread of antibiotic resistance (usually referred to TET but also to MLSB and other antibiotics) to many important pathogens. In C. difficile, the best-known element of this family is Tn5397, a 21kb element able to transfer between C. difficile and Bacillus subtilis or E. faecalis in vitro.66,67 Tn5397 differs from Tn916 for the presence of a Group II intron and for a different excision/insertion module. In fact, Tn916 contains two genes, xisTn and intTn, encoding an excisionase and a tyrosine integrase, whereas Tn5397 has a tndX gene that encodes a large serine recombinase.68 Furthermore, Tn916 inserts into multiple regions of the C. difficile genome,<sup>64</sup> while Tn5397 inserts DNA predicted filamentation processes induced by cyclic adenosine monophosphate (Fic) domain.69

Although tetM is the predominant class in *C. difficile*, other tet genes have been identified. In particular, the presences of both tetM and tetW have been described in *C. difficile* isolates

from humans and animals.<sup>70,71</sup> Furthermore, other integrative mobile genetic elements probably have a role in resistance to TET. An interesting element of 106 kb, the Tn6164, has been identified in *C. difficile* strain M120, a RT078 isolate.<sup>72</sup> This transposon is composed by parts of other elements from different bacteria, particularly from *Thermoanaerobacter* sp. and *Streptococcus pneumoniae*. Even if M120 is susceptible to TET and streptomycin, Tn6164 contains tet (44) and ant (6)-Ib predicted to confer resistance to these antibiotics, respectively.

#### Chloramphenicol (CHL)

Resistance to CHL is not so common in *C. difficile* and only 3.7% of European clinical isolates have been found resistant to this antibiotic.<sup>32</sup> *C. difficile* resistance to CHL is usually conferred by a catD gene, encoding for a CHL acetyltransferase.<sup>73,74</sup> The catD gene is located on the transposons Tn4453a and Tn4453b, structurally and functionally related to the *Clostridium perfringens* mobilizable element Tn4451.<sup>75</sup>

#### **Recurrent of the Disease**

The possibility of CDI recurrence after an initial occurrence is reported to be between 10% and 20% within 8 weeks and further increases with every other event up to 40-65%.<sup>76</sup> It is measured similar for the treatment with MTZ and VAN. Fewer secondary recurrences are also reported after treatment with fidaxomicin for patients with mild-to-moderate disease.77 The first recurrence event can be treated with the same regimens used for the initial occurrence, it all depends on the severity of disease especially in patients with high risk of additional recurrence treatment with fidaxomicin. For the second failure of CDI, MTZ is no longer option based on concerns about its side effects, especially neuropathy. In such kind of situation fidaxomicin (200 mg twice daily for 10 days) or VAN (125 mg 4 times daily for 10 days) followed by either a pulsed or tapered strategy is mostly highly recommended. McFarland et al.<sup>76</sup> were able to reduce the frequency of relapse to 14.3% by a pulsed regime and up to 31% by using a tapered strategy.<sup>78</sup> The pulsed strategy proposed a standard VAN course over 10 days followed by a course of 125 mg VAN every 2-3 days for 10 doses. Of the multiple strategies used for tapering VAN, the IDSA guidelines 2010 recommended stepping down to 125 mg twice daily for a week after the regular 10 days of VAN, followed by 125 mg once daily for a week which is then followed by pulse of 125 mg every 2-3 days for 2-8 weeks.<sup>16</sup> In case of patients with multiple recurrences, the European as well as the American guideline on CDI recommends that the intestinal microbiota transplantation (IMT) must be considered for those patients.

# The IMT

The IMT (Microbiome transfer, or fecal microbiota transplantation), is a procedure in which fecal matter is collected from a tested donor, mixed with a saline or other solution, strained, and placed in a patient, by colonoscopy, endoscopy, sigmoidoscopy, or enema, to cure the underline disease.

In early fourth century, the stool transfer treatment for diarrheal diseases was successfully practiced in China.<sup>79</sup> It was described first as a treatment option for pseudomembranous colitis in 1958.<sup>80</sup> It has been found that the intestinal microbiota in patients with CDI had a reduced bacterial diversity, as compared with healthy individuals. It has been also found that in patients with recurrent CDI, the IMT (infusion of donor feces) resulted in better treatment outcomes as compared with VAN therapy.<sup>81</sup> During the past few years, reports have shown that the treatment for recurrent CDI by endoscopically administering the feces in the duodenum, ileocolon, or by enema, the patient's cure rates were reported to increase for up to about 92%.<sup>82</sup>

Colonoscopic stool transfer was recommended on the strength of better acceptance and avoidance of bacterial contamination of the small intestine with intestinal microbes, in addition to its higher success rate. About 200 ml volume was used via the upper digestive tract.<sup>82</sup> Furthermore, for conservative application, the response was improved using 500 ml or more of suspension (80% vs. 97%).<sup>82</sup> A highly diverse protective donor flora develops within 2 weeks following stool transplantation, predominantly natural *Bacteroides* species.<sup>83</sup> To do this, a protocol-based treatment was schedule, which could be followed. This trail must be monitored for a long-term follow-up. In animal trails, a correlation between altered intestinal microbiome and the development of autoimmune diseases and obesity was observed.<sup>84</sup>

Despite its increased rate for successful curing advanced CDI cases, IMT has important limitations such as the reluctance of patients, and also physicians, to choose this treatment strategy at an early stage. Furthermore, there are legitimate concerns that IMT can spread other infectious diseases including HIV or hepatitis. There are also concerns that IMT could change the microbiome and consequently increase susceptibility to chronic conditions such as obesity or autoimmune disorders.<sup>85</sup>

# **DNA Vaccination**

DNA vaccination offers a unique platform to study the optimal antigenic regions from both toxins as this approach is able to test the immunogenicity of candidate antigens in animals directly without first producing actual antigenic proteins *in vitro*. Once high-level antibody responses are elicited, the same antigen region can be used to produce subunit-based recombinant toxin proteins as vaccines. Furthermore, the same toxin antigens can be used to produce hyperimmune sera that can be administered for passive antibody protection. Monoclonal antibodies can also be generated from a high responder host (animal or human volunteers) who received the novel N-terminal region from the *C. difficile* toxin B.<sup>86</sup>

Many efforts by different pharmaceutical companies to develop vaccines against CDI. Passive protection with antitoxin monoclonal antibodies has also been proven to be effective in reducing recurrent CDI.87 The important element for both active and passive vaccination approaches is the discovery of high quality protective antibody responses against two key C. difficile toxins.<sup>88</sup> Although, antitoxin antibodies have been widely investigated, less is known about the immunogenicity potential of toxin B. It might be the size and the highly unstable nature of both toxins, especially toxin B, has made the use of full-length recombinant protein-based vaccines less practical. While it is generally well established that the C-terminal receptor binding regions of both toxins (A and B) are ideal candidates for eliciting protective antibody responses, other areas of both toxins in eliciting protective antibody responses are yet to be investigated.

N-terminal region of toxin B protein is found to be an excellent immunogen to elicit protective antibodies, effective not only in protecting cells in an *in vitro* cytotoxicity assay but also functional in improving the protection of mice against a lethal *C. difficile* toxin when used in combination with antitoxin A antibodies. The discovery of this novel N-terminal domain of TcdB as a protective immunogen will offer more options to design the next generation subunit-based TcdB or TcdA-TcdB combination vaccines.<sup>86</sup>

The production of a panel of mAb against the toxin A of *C. difficile* elicited by DNA immunization has been described. Due to the large size of *C. difficile* toxins and the toxicity of natural materials, it is appropriate to use a part of the toxin protein as the immunogen. DNA immunization is an ideal approach for such investigations as various subdomains of the toxins can be designed and tested while there is no need for the production and purification of actual recombinant protein immunogens *in vitro*.<sup>89</sup>

Toxin A-specific mAb showed high antigen specificity and high antibody affinity. They have preserved functional activity as protective antibodies based on both *in vitro* cell protection and *in vivo* protection. More significantly, these mAb targeted different epitopes of toxin A, which allowed for the development of a "cocktail" formulation to improve the *in vivo* protein or a paired detection kit to measure the presence and levels of toxin A in testing samples, including clinical samples. One study reported a significant correlation between toxin B protein concentrations and severities of clinical CDI.<sup>90</sup>

Given the recent rapid progress in mAb technology, including the direct cloning of Ig genes from immunized B cells from both animal and human sources,<sup>91-94</sup> great opportunities to study and optimize the immunization approaches that can be effective in eliciting high titer and high affinity antibody responses in the hosts are becoming available. In combination with advanced sequencing technology, the entire process of antigen-specific B-cell development can be monitored.

## **Natural Products**

Failure rate of existing antibiotics in combating *C. difficile* seems to be high and increasing, and the recurrent infections are frequently observed. This may be attributed to the excessive and widespread use of antibiotics. Consequently, choices for *C. difficile* treatment using conventional antibiotics is becoming limited, and the development of alternative therapeutic approaches, including plant-source remedies that are usually used in traditional medicine, are undoubtedly needed to prevent and contain the spread of resistance and to ensure an effective therapy against CDI.<sup>28,95</sup>

Plant extracts were considered to be significant for various diseases by the ancient civilizations.<sup>96</sup> It is estimated that there are about 250,000 species of higher plants in the world, and pharmacological activities for most of them are yet to be investigated.<sup>97</sup> Natural products and their derivatives are the source for more than 50% of the drugs currently available worldwide, in which higher plants contribution is about 25%.<sup>98</sup> Flowering plants are producing a variety of potent drugs, for example pilocarpine to treat glaucoma and dry mouth is derived from *Pilocarpus* spp. Reserpine and other hypertensive and tranquilizing alkaloids have been isolated from *Rauwolfia* spp.<sup>99</sup> There is an international raising trend to shift resources from allopathic to traditional health-care systems.<sup>100</sup>

Microbial organisms and higher plants have been used as a natural source for the discovery of new drugs. Artemisinin, quinine, and licochalcone A are the examples for plant derived products and amphotericin B are most important antiparasitics components isolated from microorganisms. Many other natural plant products have demonstrated antiparasitic activity in the laboratory and have represent the interesting and novel structures for the development of new and immediate needed antiparasitics. The plants essential oils have demonstrated anti-inflammatory, antibacterial, antiprotozoan, and antifungal activities.<sup>100-102</sup>

In comparison with antibiotics, herbal extracts, and essential oils contain different antibacterial agents that could employ a number of inhibitory mechanisms, making it difficult for pathogens to initiate resistance.<sup>103</sup>

Antiparasitic properties of many new natural product groups have been identified with their efficacy and selectivity such as plant-derived alkaloids, terpenes, and phenolics.<sup>104</sup> Natural products have been a productive source of new bioactive compounds, allowing the discovery of therapeutic agents to treat not only infectious diseases but also cancer and other immunodeficiencies.<sup>105,106</sup> Extracts and essential oils were effective in controlling the growth of a wide variety of microorganisms, including bacteria, parasites, yeasts, and filamentous fungi.

The increased popularity of using natural product as alternative curing approach may be attributed to the belief that these products are safe and they are widely available at low costs. In traditional medicine, the use of plants in the form of crude extracts, infusions or plasters is a well-known practice to treat common infections in many parts of the world. So far, there is limited literature available regarding the natural products used against *C. difficile*, and few reports available in which the researchers have used essential oils<sup>107-109</sup> and fruits extract<sup>110</sup> against *C. difficile*. An interesting study was conducted to determine the effect of essential oil compounds on mixed fecal microbiota. The study concludes that thymol and geraniol at around 100 ppm could be effective in suppressing pathogens in the small intestine, without damaging the beneficial commensal colonic bacteria.<sup>107</sup>

Another plant product; virgin coconut oil (VCO) is found to have an antimicrobial activity. VCO active fatty acids have been tested for their antimicrobial potential against C. difficile in vitro trail. The results have shown the inhibitory effects on growth (P < 0.001) when exposed to lauric acid (C12) when it was determined by colony-forming units per milliliter. Capric acid (C10) and caprylic acid (C8) have shown lesser degree of growth inhibition. VCO could not inhibit the C. difficile growth. However, bacterial cells could not grow when exposed to 0.15-1.2% lipolyzed coconut oil. The results of transmission electron microscopy clearly showed that at 2 mg/mL of lauric acid exposure both cell membrane and cytoplasm of cells was disrupted. Furthermore, the physiological changes in bacterial cell membrane integrity were also confirmed using live/ dead staining following treatment with selected fatty acids. Inhibition of C. difficile using medium-chain fatty acids derived from VCO has been also demonstration.<sup>109</sup>

Finegold *et al.*, have investigated the effect of pomegranate extract in the management or prevention of CDIs or colonization. The activity of pomegranate was tested against 29 clinical *C. difficile* isolates using the Clinical and Laboratory Standards Institute-approved AD technique. Total phenolics content of the pomegranate extract was determined by Folin-Ciocalteau colorimetric method and final concentrations of 6.25-400  $\mu$ g/mL gallic acid equivalent were achieved in

the agar. All strains had MICs at 12.5-25 mg/mL gallic acid equivalent range. The results suggest antimicrobial activity to be a useful tool in the management and prevention of *C. difficile* disease or colonization.<sup>110</sup>

The chemical composition and the antimicrobial activity of the essential oil of *Angelica archangelica* L. (Apiaceae) roots from central Italy were analyzed. The major constituents of the oil were  $\alpha$ -pinene (21.3%),  $\delta$ -3-carene (16.5%), limonene (16.4%), and  $\alpha$ -phellandrene (8.7%). The oil shows a good antimicrobial activity against *C. difficile*, *C. perfringens*, *E. faecalis*, *Eubacterium limosum*, *Peptostreptococcus anaerobius*, and *Candida albicans* with MIC values of 0.25%, 0.25%, 0.13%, 0.25%, 2.25%, and 0.50% v/v, respectively. Interestingly, weak antimicrobial activity against the useful intestinal microflora; bifidobacteria and lactobacilli has been observed with MIC values >4.0% v/v.<sup>108</sup>

Nigella sativa known as black seeds (BSs) have been used traditionally for treating various diseases in the Middle East regions for more than 2000 years.<sup>111</sup> Different active compounds in BS have been demonstrated for their antibacterial and antifungal activities such as thymoquinone.<sup>112</sup> BS were found to inhibit growth of many Gram-positive and Gram-negative bacteria including *Staphylococcus aureus*, Pseudomonas aeruginosa, Escherichia coli, Bacillus subtilis, Klebsiella pneumoniae, Salmonella Typhimurium, and Shigella flexneri.<sup>113-116</sup> Another traditional herbal product is Commiphora myrrha (Myrrh). It has been also used in different medical contexts as astringent, antiseptic, antiparasitic, and antitussive and for treating leprosy, syphilis, and cancer.<sup>117</sup> Previous reports have indicated antibacterial activity of Myrrh against E. coli, S. aureus, B. subtilis, Bacillus circulans, E. faecalis, P. aeruginosa, and Helicobacter pylori.<sup>118-120</sup> In a recently published study, we have investigated both herbal extracts of BSs and Myrrh for their antibacterial activity against two strains of C. difficile. We have found that BS oil (2%) and Myrrh water extract are effective natural antibacterial agents to inhibit C. difficile.121 In the later study, in vitro investigation suggest that the acidic environment of the human stomach (i.e., extreme acidic environment) does not compromise the effectiveness of treating human infection with C. difficile by oral administration of BS oil (2%) and Myrrh water extract.

#### Conclusion

While IMT is considered as a promising approach with better outcome for treatment, especially in patient with recurrent CDI,<sup>81</sup> different antibiotics such as VAN, metronidazole, rifamycins and VAN are proven to have effective activity against CDI. However, overuse of such antibiotics, leading to possible emergence of antibiotic-resistant strains is potential threat. Therefore, natural antibacterial products, such as herbal extracts and essential oils that employ different inhibitory mechanisms, making it difficult for pathogens to develop resistance are potential future option. Taking into consideration that some of these natural products, including *C. myrrha* (Myrrh), could have more inhibitory activity on *C. difficile* compared with their no-observed effects on other gut normal flora.<sup>121</sup> Further investigation is needed to explore the effect of these natural products on *C. difficile* pathogenesis, including toxin gene expression and colonization, *in vivo*.

#### References

- 1. Stecher B, Hardt WD. Mechanisms controlling pathogen colonization of the gut. Curr Opin Microbiol 2011;14:82-91.
- Jin K, Wang S, Huang Z, Lu S. *Clostridium difficile* infections in China. J Biomed Res 2010;24:411-6.
- Carroll KC, Bartlett JG. Biology of *Clostridium difficile*: Implications for epidemiology and diagnosis. Annu Rev Microbiol 2011;65:501-21.
- Ananthakrishnan AN. *Clostridium difficile* infection: Epidemiology, risk factors and management. Nat Rev Gastroenterol Hepatol 2011;8:17-26.
- McDonald LC, Coignard B, Dubberke E, Song X, Horan T, Kutty PK, et al. Recommendations for surveillance of *Clostridium difficile*associated disease. Infect Control Hosp Epidemiol 2007;28:140-5.
- Khanna S, Pardi DS, Aronson SL, Kammer PP, Orenstein R, St. Sauver JL, *et al.* The epidemiology of community-acquired *Clostridium difficile* infection: A population-based study. Am J Gastroenterol 2012;107:89-95.
- Rineh A, Kelso MJ, Vatansever F, Tegos GP, Hamblin MR. *Clostridium difficile* infection: Molecular pathogenesis and novel therapeutics. Expert Rev Anti Infect Ther 2014;12:131-50.
- Voth DE, Ballard JD. *Clostridium difficile* toxins: Mechanism of action and role in disease. Clin Microbiol Rev 2005;18:247-63.
- Ho JG, Greco A, Rupnik M, Ng KK. Crystal structure of receptorbinding C-terminal repeats from *Clostridium difficile* toxin A. Proc Natl Acad Sci U S A 2005;102:18373-8.
- Aktories K, Barbieri JT. Bacterial cytotoxins: Targeting eukaryotic switches. Nat Rev Microbiol 2005;3:397-410.
- Furuya-Kanamori L, Marquess J, Yakob L, Riley TV, et al. Asymptomatic Clostridium difficile colonization: Epidemiology and clinical implications. BMC Infect Dis 2015;15:516.
- Grünewald T, Kist M, Mutters R, Ruf BR, Kern WV. *Clostridium difficile* infection. Dtsch Med Wochenschr 2010;135:699-703.
- Weis S, John E, Lippmann N, Mössner J, Lübbert C. *Clostridium difficile* infection (CDI) in the course of time An issue only for the internist? Zentralbl Chir 2014;139:460-8.
- Dansinger ML, Johnson S, Jansen PC, Opstad NL, Bettin KM, Gerding DN. Protein-losing enteropathy is associated with *Clostridium difficile* diarrhea but not with asymptomatic colonization: A prospective, case-control study. Clin Infect Dis 1996;22:932-7.
- Bauer MP, Notermans DW, van Benthem BH, Brazier JS, Wilcox MH, Rupnik M, *et al. Clostridium difficile* infection in Europe: A hospitalbased survey. Lancet 2011;377:63-73.
- Cohen SH, Gerding DN, Johnson S, Kelly CP, Loo VG, McDonald LC, et al. Clinical practice guidelines for *Clostridium difficile* infection in adults: 2010 update by the society for healthcare epidemiology of America (SHEA) and the infectious diseases society of America (IDSA). Infect Control Hosp Epidemiol 2010;31:431-55.
- Surawicz CM, Brandt LJ, Binion DG, Ananthakrishnan AN, Curry SR, Gilligan PH, *et al.* Guidelines for diagnosis, treatment, and prevention of *Clostridium difficile* infections. Am J Gastroenterol 2013;108:478-98.
- 18. Debast SB, Bauer MP, Kuijper EJ; European Society of Clinical

Microbiology and Infectious Diseases. European Society of Clinical Microbiology and Infectious Diseases: Update of the treatment guidance document for *Clostridium difficile* infection. Clin Microbiol Infect 2014;20 Suppl 2:1-26.

- Crobach MJ, Dekkers OM, Wilcox MH, Kuijper EJ. European Society of Clinical Microbiology and Infectious Diseases (ESCMID): Data review and recommendations for diagnosing *Clostridium difficile*infection (CDI). Clin Microbiol Infect 2009;15:1053-66.
- Dubberke ER, Han Z, Bobo L, Hink T, Lawrence B, Copper S, et al. Impact of clinical symptoms on interpretation of diagnostic assays for *Clostridium difficile* infections. J Clin Microbiol 2011;49:2887-93.
- Lanzas C, Dubberke ER, Lu Z, Reske KA, Grohn YT. Epidemiological model for *Clostridium difficile* transmission in healthcare settings. Infect Control Hosp Epidemiol 2011;32:553-61.
- 22. Planche TD, Davies KA, Coen PG, Finney JM, Monahan IM, Morris KA, *et al.* Differences in outcome according to *Clostridium difficile* testing method: A prospective multicentre diagnostic validation study of *C. difficile* infection. Lancet Infect Dis 2013;13:936-45.
- Stahlmann J, Schönberg M, Herrmann M, von Müller L. Detection of nosocomial *Clostridium difficile* infections with toxigenic strains despite negative toxin A and B testing on stool samples. Clin Microbiol Infect 2014;20:O590-2.
- 24. Vonberg RP, Kuijper EJ, Wilcox MH, Barbut F, Tüll P, Gastmeier P, *et al.* Infection control measures to limit the spread of *Clostridium difficile.* Clin Microbiol Infect 2008;14 Suppl 5:2-20.
- Henrich TJ, Krakower D, Bitton A, Yokoe DS. Clinical risk factors for severe *Clostridium difficile*-associated disease. Emerg Infect Dis 2009;15:415-22.
- Lungulescu OA, Cao W, Gatskevich E, Tlhabano L, Stratidis JG. CSI: A severity index for *Clostridium difficile* infection at the time of admission. J Hosp Infect 2011;79:151-4.
- Abou Chakra CN, Pepin J, Valiquette L. Prediction tools for unfavourable outcomes in *Clostridium difficile* infection: A systematic review. PLoS One 2012;7:e30258.
- Spigaglia P. Recent advances in the understanding of antibiotic resistance in *Clostridium difficile* infection. Ther Adv Infect Dis 2016;3:23-42.
- 29. Goldman P. The development of 5-nitroimidazoles for the treatment and prophylaxis of anaerobic bacterial infections. J Antimicrob Chemother 1982;10 Suppl A:23-33.
- Jarrad AM, Karoli T, Blaskovich MA, Lyras D, Cooper MA. *Clostridium difficile* drug pipeline: Challenges in discovery and development of new agents. J Med Chem 2015;58:5164-85.
- Vardakas KZ, Polyzos KA, Patouni K, Rafailidis PI, Samonis G, Falagas ME. Treatment failure and recurrence of *Clostridium difficile* infection following treatment with vancomycin or metronidazole: A systematic review of the evidence. Int J Antimicrob Agents 2012;40:1-8.
- Freeman J, Vernon J, Morris K, Nicholson S, Todhunter S, Longshaw C, et al. Pan-European longitudinal surveillance of antibiotic resistance among prevalent *Clostridium difficile* ribotypes. Clin Microbiol Infect 2015;21:248. e9-248. e16.
- Alqumber MA. *Clostridium difficile* in retail baskets, trolleys, conveyor belts, and plastic bags in Saudi Arabia. Saudi Med J 2014;35:1274-7.
- 34. Adler A, Miller-Roll T, Bradenstein R, Block C, Mendelson B, Parizade M, et al. A national survey of the molecular epidemiology of *Clostridium difficile* in Israel: The dissemination of the ribotype 027 strain with reduced susceptibility to vancomycin and metronidazole. Diagn Microbiol Infect Dis 2015;83:21-4.
- 35. Lynch T, Chong P, Zhang J, Hizon R, Du T, Graham MR, *et al.* Characterization of a stable, metronidazole-resistant *Clostridium difficile* clinical isolate. PLoS One 2013;8:e53757.

- 36. Freeman J, Stott J, Baines SD, Fawley WN, Wilcox MH. Surveillance for resistance to metronidazole and vancomycin in genotypically distinct and UK epidemic *Clostridium difficile* isolates in a large teaching hospital. J Antimicrob Chemother 2005;56:988-9.
- Moura I, Spigaglia P, Barbanti F, Mastrantonio P. Analysis of metronidazole susceptibility in different *Clostridium difficile* PCR ribotypes. J Antimicrob Chemother 2013;68:362-5.
- Baines SD, O'Connor R, Freeman J, Fawley WN, Harmanus C, Mastrantonio P, *et al.* Emergence of reduced susceptibility to metronidazole in *Clostridium difficile*. J Antimicrob Chemother 2008;62:1046-52.
- Yu X, Sun D. Macrocyclic drugs and synthetic methodologies toward macrocycles. Molecules 2013;18:6230-68.
- Perkins HR, Nieto M. The chemical basis for the action of the vancomycin group of antibiotics. Ann NY Acad Sci 1974;235:348-63.
- Brouwer MS, Warburton PJ, Roberts AP, Mullany P, Allan E. Genetic organisation, mobility and predicted functions of genes on integrated, mobile genetic elements in sequenced strains of *Clostridium difficile*. PLoS One 2011;6:e23014.
- 42. Brouwer MS, Roberts AP, Mullany P, Allan E. *In silico* analysis of sequenced strains of *Clostridium difficile* reveals a related set of conjugative transposons carrying a variety of accessory genes. Mob Genet Elements 2012;2:8-12.
- Ammam F, Marvaud JC, Lambert T. Distribution of the vanG-like gene cluster in *Clostridium difficile* clinical isolates. Can J Microbiol 2012;58:547-51.
- Ammam F, Meziane-Cherif D, Mengin-Lecreulx D, Blanot D, Patin D, Boneca IG, *et al.* The functional vanGCd cluster of *Clostridium difficile* does not confer vancomycin resistance. Mol Microbiol 2013;89:612-25.
- 45. Leeds JA, Sachdeva M, Mullin S, Barnes SW, Ruzin A. *In vitro* selection, via serial passage, of *Clostridium difficile* mutants with reduced susceptibility to fidaxomicin or vancomycin. J Antimicrob Chemother 2014;69:41-4.
- Dapa T, Leuzzi R, Ng YK, Baban ST, Adamo R, Kuehne SA, *et al.* Multiple factors modulate biofilm formation by the anaerobic pathogen *Clostridium difficile*. J Bacteriol 2013;195:545-55.
- Young GP, Ward PB, Bayley N, Gordon D, Higgins G, Trapani JA, et al. Antibiotic-associated colitis due to *Clostridium difficile*: Doubleblind comparison of vancomycin with bacitracin. Gastroenterology 1985;89:1038-45.
- Iv EC, Iii EC, Johnson DA. Clinical update for the diagnosis and treatment of *Clostridium difficile* infection. World J Gastrointest Pharmacol Ther 2014;5:1-26.
- Chaparro-Rojas F, Mullane KM. Emerging therapies for *Clostridium difficile* infection focus on fidaxomicin. Infect Drug Resist 2013;6:41-53.
- Mullane KM, Gorbach S. Fidaxomicin: First-in-class macrocyclic antibiotic. Expert Rev Anti Infect Ther 2011;9:767-77.
- Srivastava A, Talaue M, Liu S, Degen D, Ebright RY, Sineva E, *et al.* New target for inhibition of bacterial RNA polymerase: 'Switch region'. Curr Opin Microbiol 2011;14:532-43.
- 52. Miller MA, Blanchette R, Spigaglia P, Barbanti F, Mastrantonio P. Divergent rifamycin susceptibilities of *Clostridium difficile* strains in Canada and Italy and predictive accuracy of rifampin Etest for rifamycin resistance. J Clin Microbiol 2011;49:4319-21.
- Huang JS, Jiang ZD, Garey KW, Lasco T, Dupont HL. Use of rifamycin drugs and development of infection by rifamycin-resistant strains of *Clostridium difficile*. Antimicrob Agents Chemother 2013;57:2690-3.
- 54. Eitel Z, Terhes G, Sóki J, Nagy E, Urbán E. Investigation of the MICs of fidaxomicin and other antibiotics against Hungarian *Clostridium difficile* isolates. Anaerobe 2015;31:47-9.

- Terhes G, Maruyama A, Latkóczy K, Szikra L, Konkoly-Thege M, Princz G, et al. In vitro antibiotic susceptibility profile of *Clostridium* difficile excluding PCR ribotype 027 outbreak strain in Hungary. Anaerobe 2014;30:41-4.
- Curry SR, Marsh JW, Shutt KA, Muto CA, O'Leary MM, Saul MI, et al. High frequency of rifampin resistance identified in an epidemic *Clostridium difficile* clone from a large teaching hospital. Clin Infect Dis 2009;48:425-9.
- Johnson S, Schriever C, Patel U, Patel T, Hecht DW, Gerding DN. Rifaximin Redux: Treatment of recurrent *Clostridium difficile* infections with rifaximin immediately post-vancomycin treatment. Anaerobe 2009;15:290-1.
- Carman RJ, Boone JH, Grover H, Wickham KN, Chen L. *In vivo* selection of rifamycin-resistant *Clostridium difficile* during rifaximin therapy. Antimicrob Agents Chemother 2012;56:6019-20.
- Obuch-Woszczatynski P, Dubiel G, Harmanus C, Kuijper E, Duda U, Wultanska D, *et al.* Emergence of *Clostridium difficile* infection in tuberculosis patients due to a highly rifampicin-resistant PCR ribotype 046 clone in Poland. Eur J Clin Microbiol Infect Dis 2013;32:1027-30.
- Dong D, Zhang L, Chen X, Jiang C, Yu B, Wang X, et al. Antimicrobial susceptibility and resistance mechanisms of clinical *Clostridium difficile* from a Chinese tertiary hospital. Int J Antimicrob Agents 2013;41:80-4.
- Lachowicz D, Pituch H, Obuch-Woszczatynski P. Antimicrobial susceptibility patterns of *Clostridium difficile* strains belonging to different polymerase chain reaction ribotypes isolated in Poland in 2012. Anaerobe 2015;31:37-41.
- 62. Norman KN, Scott HM, Harvey RB, Norby B, Hume ME. Comparison of antimicrobial susceptibility among *Clostridium difficile* isolated from an integrated human and swine population in Texas. Foodborne Pathog Dis 2014;11:257-64.
- 63. Spigaglia P, Carucci V, Barbanti F, Mastrantonio P. ErmB determinants and Tn916-Like elements in clinical isolates of *Clostridium difficile*. Antimicrob Agents Chemother 2005;49:2550-3.
- 64. Mullany P, Williams R, Langridge GC, Turner DJ, Whalan R, Clayton C, *et al.* Behavior and target site selection of conjugative transposon Tn916 in two different strains of toxigenic *Clostridium difficile.* Appl Environ Microbiol 2012;78:2147-53.
- Dong H, Fink K, Züst R, Lim SP, Qin CF, Shi PY. Flavivirus RNA methylation. J Gen Virol 2014;95:763-78.
- Mullany P, Wilks M, Lamb I, Clayton C, Wren B, Tabaqchali S. Genetic analysis of a tetracycline resistance element from *Clostridium difficile* and its conjugal transfer to and from *Bacillus subtilis*. J Gen Microbiol 1990;136:1343-9.
- Jasni AS, Mullany P, Hussain H, Roberts AP. Demonstration of conjugative transposon (Tn5397)-mediated horizontal gene transfer between *Clostridium difficile* and *Enterococcus faecalis*. Antimicrob Agents Chemother 2010;54:4924-6.
- 68. Roberts AP, Johanesen PA, Lyras D, Mullany P, Rood JI. Comparison of Tn5397 from *Clostridium difficile*, Tn916 from *Enterococcus faecalis* and the CW459tet(M) element from *Clostridium perfringens* shows that they have similar conjugation regions but different insertion and excision modules. Microbiology 2001;147:1243-51.
- Wang H, Smith MC, Mullany P. The conjugative transposon Tn5397 has a strong preference for integration into its *Clostridium difficile* target site. J Bacteriol 2006;188:4871-8.
- Spigaglia P, Barbanti F, Mastrantonio P. Tetracycline resistance gene tet(W) in the pathogenic bacterium *Clostridium difficile*. Antimicrob Agents Chemother 2008;52:770-3.
- 71. Fry PR, Thakur S, Abley M, Gebreyes WA. Antimicrobial resistance, toxinotype, and genotypic profiling of *Clostridium difficile* isolates of

swine origin. J Clin Microbiol 2012;50:2366-72.

- 72. Corver J, Bakker D, Brouwer MS, Harmanus C, Hensgens MP, Roberts AP, *et al.* Analysis of a *Clostridium difficile* PCR ribotype 078 100 kilobase island reveals the presence of a novel transposon, Tn6164. BMC Microbiol 2012;12:130.
- Wren BW, Mullany P, Clayton C, Tabaqchali S. Molecular cloning and genetic analysis of a chloramphenicol acetyltransferase determinant from *Clostridium difficile*. Antimicrob Agents Chemother 1988;32:1213-7.
- Wren BW, Mullany P, Clayton C, Tabaqchali S. Nucleotide sequence of a chloramphenicol acetyl transferase gene from *Clostridium difficile*. Nucleic Acids Res 1989;17:4877.
- Lyras D, Storie C, Huggins AS, Crellin PK, Bannam TL, Rood JI. Chloramphenicol resistance in *Clostridium difficile* is encoded on Tn4453 transposons that are closely related to Tn4451 from *Clostridium perfringens*. Antimicrob Agents Chemother 1998;42:1563-7.
- McFarland LV, Elmer GW, Surawicz CM. Breaking the cycle: Treatment strategies for 163 cases of recurrent *Clostridium difficile* disease. Am J Gastroenterol 2002;97:1769-75.
- Cornely OA, Miller MA, Louie TJ, Crook DW, Gorbach SL. Treatment of first recurrence of *Clostridium difficile* infection: Fidaxomicin versus vancomycin. Clin Infect Dis 2012;55 Suppl 2:S154-61.
- Tedesco FJ, Gordon D, Fortson WC. Approach to patients with multiple relapses of antibiotic-associated pseudomembranous colitis. Am J Gastroenterol 1985;80:867-8.
- Zhang F, Luo W, Shi Y, Fan Z, Ji G. Should we standardize the 1,700-year-old fecal microbiota transplantation? Am J Gastroenterol 2012;107:1755.
- Eiseman B, Silen W, Bascom GS, Kauvar AJ. Fecal enema as an adjunct in the treatment of *Pseudomembranous enterocolitis*. Surgery 1958;44:854-9.
- van Nood E, Vrieze A, Nieuwdorp M, Fuentes S, Zoetendal EG, de Vos WM, *et al.* Duodenal infusion of donor feces for recurrent *Clostridium difficile*. N Engl J Med 2013;368:407-15.
- Gough E, Shaikh H, Manges AR. Systematic review of intestinal microbiota transplantation (fecal bacteriotherapy) for recurrent *Clostridium difficile* infection. Clin Infect Dis 2011;53:994-1002.
- Khoruts A, Dicksved J, Jansson JK, Sadowsky MJ. Changes in the composition of the human fecal microbiome after bacteriotherapy for recurrent *Clostridium difficile*-associated diarrhea. J Clin Gastroenterol 2010;44:354-60.
- Berer K, Mues M, Koutrolos M, Rasbi ZA, Boziki M, Johner C, *et al.* Commensal microbiota and myelin autoantigen cooperate to trigger autoimmune demyelination. Nature 2011;479:538-41.
- Smith MB, Kelly C, Alm EJ. Policy: How to regulate faecal transplants. Nature 2014;506:290-1.
- Jin K, Wang S, Zhang C, Xiao Y, Lu S, Huang Z. Protective antibody responses against *Clostridium difficile* elicited by a DNA vaccine expressing the enzymatic domain of toxin B. Hum Vaccin Immunother 2013;9:63-73.
- Lowy I, Molrine DC, Leav BA, Blair BM, Baxter R, Gerding DN, et al. Treatment with monoclonal antibodies against *Clostridium* difficile toxins. N Engl J Med 2010;362:197-205.
- Gardiner DF, Rosenberg T, Zaharatos J, Franco D, Ho DD. A DNA vaccine targeting the receptor-binding domain of *Clostridium difficile* toxin A. Vaccine 2009;27:3598-604.
- Zhang C, Jin K, Xiao Y, Cheng Y, Huang Z, Wang S, et al. Potent monoclonal antibodies against *Clostridium difficile* toxin A elicited by DNA immunization. Hum Vaccin Immunother 2013;9:2157-64.
- 90. Ryder AB, Huang Y, Li H, Zheng M, Wang X, Stratton CW, et al. Assessment of *Clostridium difficile* infections by quantitative detection

of tcdB toxin by use of a real-time cell analysis system. J Clin Microbiol 2010;48:4129-34.

- Lightwood DJ, Carrington B, Henry AJ, McKnight AJ, Crook K, Cromie K, *et al.* Antibody generation through B cell panning on antigen followed by *in situ* culture and direct RT-PCR on cells harvested en masse from antigen-positive wells. J Immunol Methods 2006;316:133-43.
- Wrammert J, Smith K, Miller J, Langley WA, Kokko K, Larsen C, et al. Rapid cloning of high-affinity human monoclonal antibodies against influenza virus. Nature 2008;453:667-71.
- Liao HX, Levesque MC, Nagel A, Dixon A, Zhang R, Walter E, et al. High-throughput isolation of immunoglobulin genes from single human B cells and expression as monoclonal antibodies. J Virol Methods 2009;158:171-9.
- Smith K, Garman L, Wrammert J, Zheng NY, Capra JD, Ahmed R, et al. Rapid generation of fully human monoclonal antibodies specific to a vaccinating antigen. Nat Protoc 2009;4:372-84.
- Brantner A, Grein E. Antibacterial activity of plant extracts used externally in traditional medicine. J Ethnopharmacol 1994;44:35-40.
- Grabley S, Thiericke R. Bioactive agents from natural sources: Trends in discovery and application. Adv Biochem Eng Biotechnol 1999;64:101-54.
- Jeevan Ram A, Bhakshu LM, Venkata Raju RR. *In vitro* antimicrobial activity of certain medicinal plants from Eastern Ghats, India, used for skin diseases. J Ethnopharmacol 2004;90:353-7.
- Cragg GM, Newman DJ. Biodiversity: A continuing source of novel drug leads. Pure Appl Chem 2005;77:7-24.
- Newman DJ, Cragg GM, Snader KM. The influence of natural products upon drug discovery. Nat Prod Rep 2000;17:215-34.
- 100. Shoaib HM, Muazzam AG, Mir A, Jung SY, Matin A. Evaluation of inhibitory potential of some selective methanolic plants extracts on biological characteristics of *Acanthamoeba castellanii* using human corneal epithelial cells *in vitro*. Parasitol Res 2013;112:1179-88.
- Rodrigues FF, Costa JG, Coutinho HD. Synergy effects of the antibiotics gentamicin and the essential oil of *Croton zehntneri*. Phytomedicine 2009;16:1052-5.
- Ajayi AO, Akintola TA. Evaluation of antibacterial activity of some medicinal plants on common enteric food-borne pathogens. Afr J Microbiol Res 2010;4:314-6.
- 103. Thompson A, Meah D, Ahmed N, Conniff-Jenkins R, Chileshe E, Phillips CO, *et al.* Comparison of the antibacterial activity of essential oils and extracts of medicinal and culinary herbs to investigate potential new treatments for irritable bowel syndrome. BMC Complement Altern Med 2013;13:338.
- Kayser O, Kiderlen AF, Croft SL. Natural products as antiparasitic drugs. Parasitol Res 2003;90 Suppl 2:S55-62.
- 105. Newman DJ, Cragg GM, Holbeck S, Sausville EA. Natural products and derivatives as leads to cell cycle pathway targets in cancer

chemotherapy. Curr Cancer Drug Targets 2002;2:279-308.

- 106. Clardy J, Walsh C. Lessons from natural molecules. Nature 2004;432:829-37.
- 107. Thapa D, Louis P, Losa R, Zweifel B, Wallace RJ. Essential oils have different effects on human pathogenic and commensal bacteria in mixed faecal fermentations compared with pure cultures. Microbiology 2015;161:441-9.
- Fraternale D, Flamini G, Ricci D. Essential oil composition and antimicrobial activity of *Angelica archangelica L*. (Apiaceae) roots. J Med Food 2014;17:1043-7.
- 109. Shilling M, Matt L, Rubin E, Visitacion MP, Haller NA, Grey SF, et al. Antimicrobial effects of virgin coconut oil and its medium-chain fatty acids on *Clostridium difficile*. J Med Food 2013;16:1079-85.
- 110. Finegold SM, Summanen PH, Corbett K, Downes J, Henning SM, Li Z. Pomegranate extract exhibits *in vitro* activity against *Clostridium difficile*. Nutrition 2014;30:1210-2.
- 111. Phillips JD. Medicinal plants. Biologist 1992;39:187-91.
- Kouidhi B, Zmantar T, Jrah H, Souiden Y, Chaieb K, Mahdouani K, et al. Antibacterial and resistance-modifying activities of thymoquinone against oral pathogens. Ann Clin Microbiol Antimicrob 2011;10:29.
- Hanafy MS, Hatem ME. Studies on the antimicrobial activity of Nigella sativa seed (black cumin). J Ethnopharmacol 1991;34:275-8.
- Arici M, Sagdic O, Gecgel U. Antibacterial effect of Turkish black cumin (*Nigella sativa* L.) oils. Grasas Aceites 2005;56:259-62.
- 115. Khan AU, Ali S, Rehman AU, Ali H, Ahmad T, Waqar M, et al. Antibacterial activity of *Nigella sativa* and *Piper nigrum*. Asian J Nat Appl Sci 2013;2:173-9.
- Shohayeb M, Halawani E. Comparative antimicrobial activity of some active constituents of *N. sativa* L. World Appl Sci J 2012;20:182-9.
- 117. Al-Harbi MM, Qureshi S, Raza M, Ahmed MM, Giangreco AB, Shah AH. Anticarcinogenic effect of *Commiphora molmol* on solid tumors induced by *Ehrlich carcinoma* cells in mice. Chemotherapy 1994;40:337-47.
- Alhussaini MS, Alghonaim MI, Ibrahim KE. An evaluation of the Antimicrobial activity of *Commiphora myrrha* Nees (Engl.) oleo-gum resins from Saudi Arabia. J Med Sci 2015:198-203.
- 119. Ghaedi M, Rezaee M, Fakhari S, Ghorashi M, Pirmohamadi H, Doost RD, *et al.* Effects of ethanol extract of *Commiphora myrrha* gum against *Helicobacter pylori* and inhibition of proliferation of the human AGS cell line *in vitro*. JATMS 2014;20:106-10.
- 120. Mohamed AA, El-Baz FK, Hegazy AK, Kord MA. Chemical composition of essential oil and *in vitro* antioxidant andantimicrobial activities of crude extracts of Commiphora myrrha resin. Ind Crops Prod 2014;57:10-6.
- 121. Aljarallah KM. Inhibition of *Clostridium difficile* by natural herbal extracts. J Taibah Univ Med Sci 2016;11:427-31.